agriculture, land reform & rural development

Department:
Agriculture, Land Reform and Rural Development
REPUBLIC OF SOUTH AFRICA

VETERINARY PROCEDURAL NOTICE NO. 19 (VPN-19)

STANDARD RELATING TO THE NATIONAL RESIDUE CONTROL PLAN

2025

Approved:

Dr Mphane Molefe
National Executive Officer – Meat Safety Act, 2000
Date: 20/02/2025

CONTENTS

PART 1: DEFINITIONS

Analysing laboratory	Means an adequately equipped laboratory staffed by technically competent personnel and accredited by the South African National Accreditation System (SANAS), as capable of performing the chemical analyses stipulated in the National Residue Control Plan (NRCP) of the Department of Agriculture (hereafter referred to as DOA).
Analyst	Means a suitably experienced and qualified chemical specialist.
Animal	Means domesticated bovine, ovine, caprine, porcine, solipeds or poultry; or farmed ostriches or crocodiles; solipeds wild game; wild cloven-hoofed game, or bees.
Approved Laboratory	Means officially approved, accredited, or registered laboratory by the Competent Authority of South Africa.
Authorised person	Means any person authorised to exercise or perform any power or duty or requested to render any service (e.g. NRCP sample collection) by the Competent Authority
Batch of animals	Means a group of animals of the same species, in the same age and time range, reared on the same farm/establishment and under the same rearing conditions.
Competent Authority	Means the national authority of South Africa, which is responsible for the organisation of official controls and other official activities (e.g., conducting NRCP) in accordance with national legislation, and any other authority to which that responsibility has been conferred.
Contaminant	Means any substance not intentionally added to food which is present in such food as a result of the production (including operations carried out in crop husbandry, animal husbandry and veterinary medicine), manufacture, processing, preparation, treatment, packaging, transport or holding of such food, or as a result of environmental contamination.
Establishment	Means an abattoir, or premises where dairy or meat products, honey or hen eggs are processed for human consumption.
Matrix	Means the material from which a sample is taken, including animal body parts, fluids, excrements, tissues, products of animal origin, animal by-products, animal feed and water.
Farmed game	Means, in relation to this VPN, ostrich, farmed crocodile or other farmed game animals
National Executive Officer	Means an officer of the competent authority designated as such by the Minister of Agriculture under the Meat Safety Act, 2000 (Act No. 40 of 2000) to implement the Act.
National Mandated Laboratory	Means a laboratory that has been authorised by the national competent authority as the laboratory for the coordination and/or testing of official samples.
Official sample	

	Means a sample taken by the Competent Authority and or officially authorised person with applicable information, for the purposes of examination of the residues or substances listed in Annex G.
Provincial Executive Officer	Means an officer of the province designated as such by the Member of Executive Council of the province responsible for Agriculture to implement the Meat Safety Act, 2000 in the province.
Residue	Means a residue of substances having a pharmacological action, metabolites of such substances, degradation products of such substances, reaction products of such substances, and other related substances present in or on a food product.
Residue control plans for pharmacologically active substances, pesticides, and contaminants	Means a control plan on the use of pharmacologically active substances, the maximum residue limits of pharmacologically active substances, the maximum residue levels of pesticides and the maximum levels of contaminants in food-producing animals, products of animal origin, including those used in composite products.
Wild game	Means, in relation to this VPN, zebra, springbuck, blesbuck or other game animals that are not domesticated

PART 2: PURPOSE AND SCOPE

2.1. Purpose

The purpose of the National Residue Control Plan (NRCP) for food-producing animals and products of animal origin is to ensure effective monitoring and determination of whether export-registered establishments use prohibited or unauthorised pharmacologically active substances in the treatment and management of animals and/or their products used as food for human consumption. The NRCP also provides information as to whether the withdrawal period for a particular veterinary medicinal product is observed in the treatment of animals. The presence of pharmacologically active substances authorised as veterinary medicinal products or as feed additives and prohibited or unauthorised pharmacologically active substances and residues thereof, as well as pesticide residues or contaminants in food, may pose a risk factor for public health. This Veterinary Procedural Notice (VPN) prescribes the Standard relating to the NRCP for the control of the residues of these substances in the food chain.

Routine random sampling of animal products is done to survey for the residues of veterinary drugs/medicinal products, environmental contaminants, and agricultural compounds and pesticides to determine if additional control measures by the Government are required. However, new drugs and chemicals regularly come onto the market, and the popularity of remedies increases or diminishes continuously. The occurrence of environmental sources of contamination also changes with new industries developing all the time. In the context of safety, there is a risk to human health from residues of chemical substances in food, as well as the issue of non-compliance with regulations related to residues in food due to improper farm management or agricultural practices.

Considering the above, a targeted sampling strategy is necessary, and the NRCP is updated and revised every year. Therefore, all samples must be targeted according to the criteria laid down in the NRCP.

The national risk-based control plans for production must control combinations of substance groups and commodity groups in accordance with the annually updated NRCP. The sample taken is considered an official sample, and it is collected from food-producing animals and animal products originating from the territory of South Africa, not from imports.

The annually adopted and implemented NRCP provides for the monitoring of certain substances and their residues in food-producing animals and animal-origin products and establishes sampling levels and sampling frequencies, as well as groups of substances to be monitored for each food commodity listed and intended to be listed for the export to European Union (EU) and other applicable markets.

The NRCP is targeted to consider the following minimum criteria: species, gender, age, production seasonality, fattening system, all available background information, and all evidence of misuse or illegal use of substances. Additionally, suspect samples are taken as part of residue control.

A risk-based assessment study must be conducted each year to determine which pharmacologically active substances, pesticides, and contaminants must be singled out for monitoring and control in accordance with the criteria listed in Paragraph 8. The results obtained for the previous year's NRCP must also be considered. Additionally, the results of food business operators' controls and the outcomes of other official controls (animal health and welfare, food safety, etc.) must be considered.

Therefore, the NRCP for pharmacologically active substances, pesticides and contaminants must provide guarantees as regards compliance with the EU and other applicable markets' export requirements on the following:

i) the use of pharmacologically active substances;
ii) maximum residue limits of pharmacologically active substances;
iii) maximum residue levels of pesticides; and
iv) maximum levels of contaminants.

2.2. Scope

South Africa is required to implement a residue monitoring plan for veterinary drugs and prohibited substances in food products of animal origin. When South Africa wishes to export animal products to the EU and other markets, it is required to satisfy the European Commission that its legislation, controls, and residue surveillance measures provide equivalent guarantees for consumers. The scope of testing under the NRCP is very comprehensive and risk-based, covering various food groups and four broad residue categories: banned substances, such as growth-promoting hormones and growth-promoting hormones with antimicrobial activity; authorised veterinary medicines; approved animal feed additives and environmental contaminants. This monitoring program helps protect consumers by ensuring high compliance with domestic and export market regulations.

The provisions of the EU legislation and Directorate-General for Health and Food Safety (DG SANTE) Guidelines on EU requirements for entry of animals and products of animal origin and Control plans for residues of veterinary medicines, pesticides, and contaminants for third countries are used as the basis for the procedure.

PART 3: RESPONSIBILITIES

3.1. Authorised person

The national executive officer must authorise competent persons to collect and store the samples and to organise the transport of the official samples to the mandated or any other approved laboratory under appropriate conditions.

3.2. Approved establishment

An establishment approved by the Competent Authority to export fresh meat, processed meat products, honey, dairy products, or hens' eggs to countries that require monitoring such products for residues must be registered by the Competent Authority.

Please refer to VPN/01 for the procedures to register an establishment for export.

3.3. Approved farm

Any farm that produces animals for export slaughter or farms that produce honey, raw milk, or hens' eggs for producing products intended for export must be registered by an Authorised person.

Please refer to VPN/02, VPN/07 and VPN/44 for the procedures to register a farm.

3.4. Laboratories

The analysis of samples must be carried out exclusively by laboratories approved and or registered for official residue control by the national executive officer.

PART 4: DEVELOPING A SAMPLING PLAN

4.1. General

This VPN establishes the responsibilities, documentation, and procedures when collecting samples for the NRCP for food-producing animals and products of animal origin and the procedure in case of suspect animals and non-compliant findings. That also applies to sampling live animals' excretions and body fluids; excrements, body fluids and tissues/organs from slaughtered animals; products of animal origin (eggs, milk, and honey); feeding stuff and drinking water for animals.

Refer to Annex F: The standard operating procedure for compiling the NRCP applicable to approved export farms and establishments, attached hereto.

4.2. Substances tested according to NRCP

After approval of the NRCP for the specific commodities by the DG SANTE and listing for export to the EU and other applicable markets, practical implementation is carried out in accordance with the procedures prescribed in this VPN.

The list of substances to be tested is contained in Annex G of this VPN and listed in the templates for the specific commodities (Microsoft Excel files) that must be provided by the national executive officer, including templates for pesticides and contaminants. The templates also contain general rules and explanations about drafting plans.

4.3. Sampling plan

The sampling plan must be developed by the national executive officer and submitted electronically (by e-mail) to the Provincial Executive Officer (PEO) in each Province. The PEO must distribute the plan to state veterinary offices in the areas where the establishments/farms where the sampling must be carried out are located.

The plans must be delivered before the beginning of each year. The sampling plan must contain the following data:

a. Name and address of the province
b. Number, name, and place of the establishments/farms where sampling is to be carried out
c. Month of the year (number)/date

d. Total number of samples from animals/products and
e. Matrix.

If the sampling cannot be carried out (the establishment/farm is no longer working/operating, or the requested animals were not slaughtered in that week, etc.), the authorised person is obliged to consult with the provincial executive officer or decide on sampling in an alternative establishment/farm.

The national executive officer must evaluate the implementation of the plans every three months and deliver the report to the provincial executive officers.

4.4. Sampling strategy

Sampling must be carried out in variable intervals spread evenly over all months of the year or the relevant production period. In this context, it must be considered that a number of pharmacologically active substances are administered only in particular seasons.

Sampling must be performed at or close to slaughter, collection, or harvest. However, for Group A substances, sampling should also be performed at any relevant stage in the life cycle of the animals. All samples must be targeted according to the criteria laid down in the NRCP.

For Group A substances, sampling must target detecting illegal treatment with prohibited or unauthorised substances; thus, animals that most likely have been treated are preferentially selected over animals that have not been treated. As much of this sampling is conducted on farms, drinking water and feed samples may be appropriate in addition to inedible materials such as blood, urine, faeces, hair, etc.

For Group B substances, samples must comprise only edible tissues/products (the objective is to verify compliance with maximum residue limits (MRLs) and maximum levels (MLs)). Sampling must be targeted on products from animals that are most likely to have been treated with a specific pharmacologically active substance or substance within a therapeutic class of veterinary medicinal products.

Samples from drug injection sites can be appropriate to control the illegal use of substances. In case samples are taken from injection sites, this must be clearly mentioned when reporting analytical results from these samples.

Selection of the animals or products to be sampled for each food business operator consideration must be given to the following:

a. History of non-compliance of the farm/establishment or producer.
b. Shortcomings in the application of veterinary medicinal products, deficiencies identified in previous controls, reported an increase in losses of animals on the farm, animal health status of the farm, and epidemiological status of the region.
c. Information on the farming system, fattening system, breed, and sex of the animals.
d. Common practices with regard to the administration of particular pharmacologically active substances in the respective farm or production system.
e. Indications of the use of pharmacologically active substances.
f. The absence or the unreliability of own checks, the membership of quality assurance schemes or industry associations (when available) and testing results under such schemes.
g. Evidence of absence or insufficient supervision of the farm by veterinarians
h. Representative sampling regardless of the size of the food business operator.

Criteria for the selection of establishment: abattoir, cutting plants, establishments for milk production, establishments for honey and egg and egg packing centres from which samples should be taken, and in addition, for:

a. Group A substances – criteria for selecting specific substances for testing within each substance group:

- Frequency of the detection of non-compliance in the country or reported in the results from other countries' samples, especially when reported under the Rapid Alert System for Food and Feed (RASFF) or the Administrative Assistance and Cooperation System (AAC) in the case of the EU.
- Availability of suitable laboratory methods and analytical standards.
- Pharmacologically active substances which are likely to be misused to increase production or increase feed conversion efficiency.
- Prohibited or unauthorised substances for which there are indications of misuse.
- Possible risks for consumers or certain population groups arising from the consumption of residues present in food.

b. Group B substances - Criteria for selecting specific substances for testing within each substance group:

- Frequency of the detection of non-compliance in the NRCP samples or other countries' samples, especially when reported under the RASFF.
- Availability of suitable laboratory methods and analytical standards.
- Information on the quantities of veterinary medicinal products produced, imported, exported, marketed, and sold for a specific food-producing animal species.
- Information on the veterinary medicinal product distribution chain, the national register of pharmacologically active substances authorised as veterinary medicinal products or feed additives, and information on the most popular prescribing patterns.
- The likelihood of misuse of the pharmacologically active substances.
- MRLs and MLs for pharmacologically active substances and feed additives including restrictions (e.g. not for use in lactating animals).
- Formulations of veterinary medicinal products for which long withdrawal periods, post-animal treatment, have been established to ensure that edible unprocessed animal products comply with EU and or other applicable markets MRLs depending on the destination of the products.
- Possible treatment of food-producing animals with veterinary drugs/medicinal products outside the terms of the marketing authorisation in food-producing terrestrial animal species (off-label use).
- The respective establishments' share of the country's total production volume.
- Non-compliances identified in earlier controls on the use of pharmacologically active substances and residues thereof.
- Origins and transport routes of the slaughtered animals, milk, eggs, or honey.
- The scope and results of own checks for residues.

4.5. General guidance for determining the matrix, sampling size and number of samples

The table below must be used as a general guideline for determining the matrix, sampling site, and minimum sample quantity per commodity (animal/food product).

Commodity	Matrix	Sampling site	Minimum sample
• Wild game • Farmed game • Cattle/pigs/sheep/goats/horses	Muscle	Slaughter line – after evisceration inspection point	500g
• Wild game* • Farmed game	Blood	On-farm	2x7 ml or 1x10 ml serum from 1 bird
• Wild game • Farmed game • Cattle/pigs/sheep/goats/ horses	Liver (without gall-bladder)	Slaughter line – evisceration inspection point	500 g
• Wild game • Farmed game			Both kidneys & kidney fat or 150g (pooled

	Kidney	Slaughter line – evisceration inspection point	sample)
Pigs			One whole kidney
Cattle/horses			One kidney or portion of the kidney (at least 100g taken from one pole)
Sheep/goats			Both kidneys or 150g (pooled sample)
Cattle/pigs/sheep/ goats/horses	Urine	• On-farm • Slaughter line - after evisceration	100 ml
Cattle/pigs/sheep/ goats/horses	Blood	• On-farm • Slaughter line – during bleeding	30 ml 30 ml
Cattle/pigs/sheep/goats/horses	Thyroid gland	Slaughter line - after evisceration	Whole gland (with surrounding tissue)
Cattle/pigs/sheep/goats/horses	Fat tissue	Slaughter line – evisceration inspection point	200 g
Poultry	Blood	Shortly after neck cutting point	30 ml**
Poultry - chickens broiler	Liver	Taken offline to enable the liver to be cut off	50 g liver tissue pooled from at least 6 birds***
Poultry - chickens broiler	Muscle (breast muscle)	Slaughter line – evisceration inspection point	100 g muscle pooled from at least 6 birds
All	Feed	On-farm	500 g
Poultry eggs	Whole Egg	On the farm or in an egg collection, sorting, and packaging centre	12 eggs
• Cow milk • Milk from other species	Raw milk	• Row milk cooling tank on farm • Transportation tank/reception site	500 ml
Honeybees	Honey	Any point of the production chain – production and packaging to ensure traceability to the beekeeper)	500 g

* Wild game blood/serum will not be collected in the field unless instructed otherwise or if there is suspicion that warrants sampling
**Collective sample from at least 25 birds from the same flock/batch of animals
*** Collective liver tissue of broilers originating from the same flock/batch of animals

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4.6. Equipment for sampling, packing and dispatching samples to the laboratory

Where the provincial competent authority has requested assistance due to a lack of resources, equipment for taking samples within the NRCP is to be procured by the national competent authority and distributed to the respective provincial offices and subsequently to authorised persons taking samples on-site.

Equipment for sampling, packaging, and dispatch of samples to the laboratory includes:

a. Disposable gloves
b. Knives
c. Plastic spoons
d. Needles for taking blood samples
e. Gel tubes/vacutainer blood collection tubes containing anticoagulant (different sizes)
f. Catheters for taking urine samples
g. Regular plastic and leak-proof containers (100 ml, 500 ml, 1000 ml)
h. Plastic safety-secure bags/envelopes (smaller for individual samples)
i. Tamper-proof safety-secure bags (bigger of different volumes, for collective samples)
j. Waterproof markers
k. Self-adhesive labels
l. Self-adhesive tape

Sampling and sample packaging equipment must be made of chemically neutral material adequate for packaging and transporting samples.

If the sample taken needs to be treated before packaging and dispatch for testing, the treatment is carried out in a clean and safe place to prevent contamination of the sample.

PART 5: SAMPLING PROCEDURE

5.1. General

Whenever official samples are taken, sampling must be unforeseen, unexpected, and effected at no fixed time and no particular day of the week. The authorised person shall perform official controls regularly, with appropriate frequencies determined on a risk basis, and must take all the precautions necessary to ensure that the element of surprise in the checks is constantly maintained. Official controls shall be performed without prior notice to the farmer/establishment (food business operator).

Sampling must be carried out in variable intervals spread over the whole year or, where applicable, over the entire harvesting/fattening/production season. In this context, it has to be considered that a number of veterinary medicinal products and/or pesticides are administered only in particular seasons.

Other available information must be considered when choosing the samples, e.g., the use of presently unknown veterinary medicinal products or pesticides, diseases suddenly appearing in particular regions, indications of fraudulent activities, and other prescribed in the sampling strategy (see section 4.4.). The comments column on the sample submission forms must include applicable information regarding possible risks.

When collecting the samples, efforts must be made to avoid multiple sampling, for example, the taking of several different samples from a single animal/product (unless the different samples are analysed for a different group of substances) or sampling several animals/products from a single producer on a given day when samples could be drawn from animals/products from several producers which would satisfy the targeting criteria, unless the operator has been identified based on the criteria included in point 8.6 or an appropriate justification has been provided in the control plan. Compliance with the planned frequency of checks must be ensured.

Each sample collected for NRCP must be accompanied by an accurately completed sample submission form (Annexes A, B(1) and B(2)) and sample dispatch form (Annexes C1, C2 and C3).

5.2. Targeted samples

Sampling for NRCP implementation must be targeted. Targeted samples are taken to detect illegal treatment or control compliance with the maximum levels in the relevant legislation. This means that the NRCP targets the groups of animals (species, gender, age) where the probability of finding residues is the highest. Conversely, the objective of random sampling is to collect significant data to evaluate, for example, consumer exposure to a specific substance.

In addition, the following should also be taken into consideration:

a. For substances listed in group A in Annex G, the objective of monitoring is to detect the use of prohibited or unauthorized substances in food-producing animals' treatments or as feed additives, that is, illegal use.
b. For the substances listed in group B in Annex G, the objective of monitoring is to control the use of authorised food-producing animals' treatments and their compliance with the maximum residue limits of veterinary drugs and maximum levels of pesticides and contaminants.

5.3. Random samples

Random samples are taken from the production lot/batch of animals, for example, when sampling broiler chickens and collecting samples of eggs, honey, and animal feed. The random sample must

be, at the same time, a representative sample of the production lot/batch quantity from which it was taken.

5.4. Suspect samples

Suspect samples are taken as a consequence of:

a. non-compliant results on samples collected in accordance with the NRCP,

b. possession or presence of prohibited substances at any point during manufacture, storage, distribution or sale through the food and feed production chain,

c. suspicion or evidence of illegal treatment or non-compliance with the withdrawal period for an authorised medicinal veterinary product or

d. other perceived reason or evidence indicating suspicion.

Note: Suspect samples taken during the follow-up of the non-compliance actions must not be counted to achieve the prescribed sampling frequency for the risk based NRCP.

PART 6: COLLECTION OF SAMPLES FOR SPECIFIC PRODUCTS

Samples must be collected as indicated in the applicable annexure

6.1. Wild game samples (see annexure H)

Wild game are animals that are hunted and shot in the wild for human consumption. Samples must be collected from all wild game animals approved/listed for export to the EU (i.e. zebra, crocodile) in accordance with the NRCP.

Blood/serum samples of wild game animals must not be collected as a standard, unless an instruction from the national competent authority has been issued to collect the samples, or there is otherwise an informed rationale for the collection of such samples by the authorised person and the information is relayed to the competent authority.

Wild game samples must be collected in the field or in abattoirs or cutting or processing establishments, where it is easier to identify the animal and the organs belonging to each individual animal.

Identification and records of every wild game animal shot must contain information that will enable traceability to where the animal was shot. In order to correctly interpret the test results, if possible, the age (estimated) and subtype of the wild game should be provided.

When collecting samples from wild game animals (in terms of matrix, sampling site and minimum quantity), the general guidance in the table in Part.4.5 above applies.

Kidney, kidney fat, liver, and muscle samples must be taken from game carcasses that have passed official control and been declared suitable for human consumption.

6.2. Farmed game samples (see annexure H)

Farmed game refers to farmed ratites (e.g., ostrich) and farmed land mammals other than domestic bovine (including Bubalus and bison species), porcine, ovine and caprine animals and domestic solipeds (mammals with a single hoof on each foot, e.g., horse).

Game reared in this way can be treated with veterinary medicinal products and other substances (unauthorized, forbidden and/or authorised). Samples of feed, liver, kidney, fat tissue and muscle tissue must be collected in accordance with the NRCP provisions.

6.3. Poultry samples (see annexure I)

When collecting samples from poultry (in terms of matrix, sampling site and minimum quantity), the general guidance in the table in Part.4.5 above applies.

Blood, feed, and drinking water samples for poultry must be collected on poultry farms.

Liver and muscle must be collected at poultry abattoirs.

Samples of liver, muscle and fat tissue must be collected from identified birds that have been inspected and from which meat and organs have been declared safe for human consumption.

Liver, muscle, and fat tissue samples should be collected at the place where the internal organs are separated (carcass evisceration site)

6.4. Bovine, sheep, goats, pigs, and horse samples

Blood and urine samples and feed and drinking water samples must be collected on farms.

Kidney, kidney fat tissue, liver, muscle tissue, thyroid gland, blood, and urine samples must be collected at abattoirs.

Samples must be collected only from identified animals that have been inspected (slaughter line control) and whose carcasses have been declared suitable for human consumption. They must be collected in accordance with this VPN.

6.5. Milk samples

6.5.1 Sampling from cows

For NRCP, milk samples must be collected from cows. Samples must be collected from farms/establishments supplying milk to dairy plants registered/approved for export.

6.5.2 Sampling from collection tank

Samples must be collected from the farm's collection tank, the tank at the collection point (if applicable), or the tank of the transport vehicle at the point of receipt of raw milk at the dairy before the tank is unloaded.

Before taking the sample, the raw milk should be lightly mixed, and the sample should be collected by dipping a special plastic container with a handle.

6.5.3 Labelling/marking samples

Each milk sample should be precisely and accurately labelled/marked and accompanied by information on farm details (farm ID (if applicable), location, address), collection point and/or transport vehicle details (collection point address, registration number of transport vehicle, serial number or mark of the vehicle tank, details of all farms from which milk was collected into the collection point or the vehicle tank. Such detailed information is essential (especially for the pooled milk sample) if the non-compliant result is received to enable tracing back to the farm of origin.

6.6. Egg samples

Egg samples must be collected at practically suitable places such as farms, collection centres and egg sorting and packing centres. All eggs samples must be traceable to the farm and house of origin.

Fresh eggs are collected only from production batches that have been tested and declared suitable for human consumption.

6.7. Honey samples

Honey samples must be collected directly from individual beekeepers during honey harvesting or, if not possible, at honey processing and packaging establishments.

When a honey sample is collected in a honey processing and packaging establishment, it should be taken from one production batch whose origin can be determined and traced back to the apiary. Therefore, when planning the collection of samples, the season of honey harvesting in different areas must be considered.

PART 7: AUDITING

Each province must be audited by the national executive officer every six months and more frequently if required, and relevant reports must be produced and provided where applicable.

PART 8: SAMPLE SUBMISSION AND REGISTRATION

Unless instructed otherwise, the samples must be submitted to the national mandated laboratory (NML) or EU accredited laboratory. Details of the NML or EU accredited laboratory are to be provided by the national competent authority.

PART 9: TRANSPORT AND STORAGE

Courier costs of NRCP samples transported to the NML or any other location under the instruction of the NEO or PEO, will be covered by the exporting food business. The courier service and transportation of samples must be approved by the NEO.

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ANNEX A
NATIONAL RESIDUE CONTROL PLAN
SAMPLE SUBMISSION FORM SAMPLES COLLECTED AT ESTABLISHMENTS

Sample registration number:

1. ESTABLISHMENT DETAILS

Establishment name:		Establishment number:	
State Veterinary Office:		Province:	
Address of establishment:			

2. SAMPLING REPORT: TO BE COMPLETED BY AUTHORIZED PERSON

1) Description of sample

Mark with X	Species	Matrix	Minimum sample size	Single sample/ Pooled sample[1] (S or P)	Identification (if known)	Analysis reference number as indicated on the sample grid
	Poultry[2]	Liver	100 g			
	Poultry	Muscle	100g muscle pooled from at least 6 birds			
	Poultry	Fat Tissue	150 g			
	Poultry	Blood	30 ml			
	Cattle	Raw milk	500 ml			
	Cattle	Muscle	500 g (For farmed or wild game samples must be at least more than 150g)			
	Wild Game[3]					
	Farmed Game[4]					
	Cattle	Blood	30 ml			
	Wild Game[5]	Blood	2x7 ml or 1x10 ml serum from 1 ostrich			
	Farmed Game					
	Cattle	Fat Tissue	200 g (For farmed or wild game samples must be at least more than 150 g)			
	Wild game					
	Farmed game					
	Cattle	Thyroid gland	Whole gland			
	Cattle	Urine	100 ml			
	Cattle	Kidney	100 g			
	Wild Game	Kidney	Both kidney & kidney fat or 150 g (pooled samples)			
	Farmed Game					

	Cattle	Liver	500 g (For farmed game and wild game sample)			
	Wild Game					
	Farmed Game					
	Honeybees	Honey	500 g			
	Poultry	Eggs	12 eggs			

(1) Samples of fat/liver/muscle obtained from poultry, wild game and farmed game from the same herd/flock may be pooled to ensure that the required minimum sample size is obtained. It must be clearly stated on this form if this was necessary.
(2) Poultry refers to chicken.
(3) Wild game refers to zebra, springbuck, blesbuck or other wild game animals.
(4) Farmed game refers to ostrich, farmed crocodile or other farmed game animals.
(5) Wild game blood/serum must not be collected in the field unless instructed otherwise or if there is suspicion that warrants sampling

2) Origin of animal(s)

a) Owner or the person having charge of the animals:

Name: _____

Address: _____

Telephone number: _____E-mail address:_____

b) Details of the farm of origin of the animal(s):

Name of farm: _____Registration number: _____

Address: _____

State Veterinary Area: _____ Province: _____

3. DECLARATIONS:

1) Declaration by authorised person

I _____(full name), hereby declare that I collected the sample/s and that the information provided in this form is accurate.

Signature: _____ Date: _____

2) Declaration by the owner or representative of the establishment

I _____(full name), hereby declare that the sample/s were collected by the authorized person mentioned above and that no relevant information was withheld from the authorized person.

Signature: _____ Date: _____

4. IDENTIFICATION OF AUTHORIZED PERSON

Name: _____ Designation: _____

Address: _____

Telephone number: _____ Email address: _____

OFFICIAL STAMP

FOR USE BY THE NML OR APPROVED LABORATORY ONLY

Substance or substances group examination: _____

Laboratory to carry out examination: _____

Date dispatched to approved laboratory: _____

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ANNEX B (1)
NATIONAL RESIDUE CONTROL PLAN
SAMPLE SUBMISSION FORM FOR ON-FARM BLOOD SAMPLES

Sample registration number: _____

1. FARM DETAILS

Farm name:		Farm registration number:	
State Veterinary Office:		Province:	
Address of farm:			

2. SAMPLING REPORT: TO BE COMPLETED BY AUTHORISED PERSON

1) Description of sample:

Type	Minimum sample size
Blood	Ostrich 2 samples (2 x 7 ml or 1 x 10 ml constitutes 1 sample), Poultry 30ml

2) Animal species:

Type:	Sex (if known)	Age (Months)	Tag number/ cycle number	Sample number/house number	Sample Reference – to be completed by NML	Comments
Wild game*						
Farmed game						
Poultry**						
Beef cattle						
Dairy cattle						

* Wild game blood/serum must not be collected in the field unless instructed otherwise or if there is suspicion that warrants sampling.

** In case of poultry, a pooled sample from at least 25 birds from the same flock/batch of animals must be collected

3) Owner or person having charge of the animals:

Name: _____

Address: _____

_____

Telephone number: _____

3. DECLARATIONS

1) Declaration by authorised person

I _____(full name), hereby declare that I collected the sample/s and that the information provided in this form is accurate.

Signature: _____ Date: _____

2) Declaration by the owner or representative of the farm

I _____(full name), hereby declare that the sample/s were collected by the authorized person mentioned above and that no relevant information was withheld from the authorized person.

Signature: _____ Date: _____

4. IDENTIFICATION OF AUTHORIZED PERSON

Name: _____ Designation: _____

Address: _____

Telephone number: _____ Email address: _____

OFFICIAL STAMP

FOR USE BY THE NML OR APPROVED LABORATORY ONLY

Substance or substances group examination: _____

Laboratory to carry out examination: _____

Date dispatched to approved laboratory: _____

agriculture, land reform & rural development
Department:
Agriculture, Land Reform and Rural Development
REPUBLIC OF SOUTH AFRICA

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ANNEX B (2)
NATIONAL RESIDUE CONTROL PLAN
SAMPLE SUBMISSION FORM FOR ON-FARM FEED AND WATER SAMPLES

Sample registration number:

1. FARM DETAILS

Farm name:		Farm registration number:	
State Veterinary Office:		Province:	
Address of farm:			

2. SAMPLING REPORT: TO BE COMPLETED BY AUTHORISED PERSON

(1) Sampling size:

SAMPLED PORTION [1]	AGGREGATE SAMPLES [2]	REDUCED SAMPLES [3]	FINAL SAMPLES [4]	Tick where appropriate
Feed trough (Loose feed) <2.5 metric tons	8 incremental samples of 500 g each to make up a total of 4 kg	Through the mixing of incremental samples	500 g	
Feed trough (Loose feed) >2.5 metric tons	Number[5] of samples in equal amounts to make up an aggregate sample of 4 kg. 2.5-10t – 10 samples 10-15t – 15 samples 15-20t – 20 samples 20-30t – 25 samples 30-40t – 30 samples >40t – 40 samples	Through the mixing of incremental samples	500 g	
Feed containers (1-4 bags)	All containers sampled in equal amounts to make up a total of 4 kg	Through the mixing of incremental samples	500 g	
Feed bags (5-16 bags)	Four bags sampled at 1 kg each should make up a total of 4 kg	Through the mixing of incremental samples	500 g	
Feed bags (more than 16 bags)	Number of samples = square root of (no of bags in sampled portion) in equal portions per bag, making up 4 kg	Through the mixing of incremental samples	500 g	
Bulk feed bin (Loose feed) <2.5 metric tons	8 incremental samples of 500 g each to make up a total of 4 kg	Through the mixing of incremental samples	500 g	
Bulk feed bin (Loose feed) >2.5 metric tons	Number of samples in equal amounts to make up an aggregate sample of 4 kg.	Through the mixing of incremental samples		

	>2.5-5t – 10 samples >5-10t – 10 samples >10-15t – 15 samples >15-20t – 20 samples >20-30t – 25 samples >30<40t – 30 samples >40t – 40 samples			
Drinking water	500 ml must be collected, and the sample must be stored at a temperature of +2 to +7˚C before delivery/dispatch to the laboratory.		500 ml	

[1] Sampled portion refer to the total amount of feed present (in a feed trough, camp, collection of feed bags or bulk feed bin) that will be sampled and is homogenous.
[2] Aggregate sample refers to the representative sample comprising a number of smaller samples (called incremental samples) obtained from the sampled portion by drawing various samples.
[3] Reduced sample – means the aggregate sample after it has been thoroughly mixed into one homogenous sample.
[4] Final sample – a final sample of at least 500 g is collected from the reduced sample.
[5] Number of samples = the square root of [tons of feed sampled x 20]. Rounded the calculated sample numbers to practical values. Lower feed weights were slightly rounded up to ensure sufficient samples are collected.

Lumps must be broken up or removed from the aggregate sample. Moist samples are to be kept refrigerated to avoid spoilage. No whole grain or roughage samples are to be collected.

In practice, this means the authorized person will identify the feed to be sampled. Depending on how much it is and in what format it is (sampled portion), he/she will collect a number of smaller samples (incremental samples) from various representative places to make up a composite sample (aggregate sample). He/she will mix the composite sample thoroughly (reduced sample) before collecting a final 500 g sample (final sample). The final sample will be sent to the laboratory, while the composite sample will be returned to the owner. The number of smaller samples (incremental samples) to be collected from the feed to be sampled (sample portion) is indicated in the table above.

(2) Identification of the sample

Type of feed sample: _____

Collection location of water sample: _____

Commercial feed or mixed on farm: _____

Name of commercial feed: _____

Batch number(s) of commercial feed: _____

In case of farm mixes, commercial name(s) of premix(es) used: _____

_____

Batch number(s) of commercial premixes: _____

Comments: _____

_____

(3) Owner or person responsible for the farm

Name: _____

Postal Address: _____

Tel number: _____ Email address: _____

3. DECLARATIONS

(1) Declaration by authorised person

I _____(full name), hereby declare that I collected the sample/s and that the information provided in this form is accurate.

Signature: _____ Date: _____

(2) Declaration by the owner or representative of the farm

I _____(full name), hereby declare that the sample/s were collected by the authorized person mentioned above and that no relevant information was withheld from the authorized person.

Signature: _____ Date: _____

4. IDENTIFICATION OF AUTHORIZED PERSON

Name: _____ Designation: _____

Address: _____

Telephone number: _____ Email address: _____

OFFICIAL STAMP

FOR USE BY THE NML OR APPROVED LABORATORY ONLY

Substance or substances group examination: _____

Laboratory to carry out examination: _____

Date dispatched to approved laboratory: _____

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ANNEX C (1)
NATIONAL RESIDUE CONTROL PLAN
DISPATCH FORM FOR SAMPLES COLLECTED AT THE ESTABLISHMENT

Establishment Name: _____ **ZA number**:_____

1. Sample description

Matrix	Analysis reference number(s)	Total number of samples dispatched
Muscle		
Fat		
Kidney		
Liver		
Blood		
Eggs		
Raw milk		

2. Details of sender – authorized person

Name: _____

Designation: _____

Address: _____

_____

Telephone number: _____

email address: _____

Official Stamp

Comments: _____

_____

Signature: _____ Dispatch date: _____

FOR USE BY THE NML OR APPROVED LABORATORY ONLY

Date received: _____ Condition of sample(s) and comments: _____

_____

Signature: _____ Date: _____

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ANNEX C (2)
NATIONAL RESIDUE CONTROL PLAN
DISPATCH FORM FOR ON-FARM FEED AND WATER SAMPLES

Farm Name: _____ **Registration number**:_____

1. Sample description

Farm names and registration numbers	Analysis Reference number(s)	Total number of samples dispatched

2. Details of sender – authorized person

Name: _____

Designation: _____

Address: _____

_____

Telephone number: _____

email address: _____

OFFICIAL STAMP

Comments: _____

_____

Signature: _____ Dispatch date: _____

FOR USE BY THE NML OR APPROVED LABORATORY ONLY

Date received:_____ Condition of sample(s) and comments: _____

_____

Signature: _____ Date: _____

agriculture, land reform & rural development
Department:
Agriculture, Land Reform and Rural Development
REPUBLIC OF SOUTH AFRICA

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ANNEX C (3)
NATIONAL RESIDUE CONTROL PLAN
DISPATCH FORM FOR ON-FARM BLOOD SAMPLES

Farm Name: _____**Registration number**:_____

Animal species:

Animal Species	**Total number of samples dispatched**	**Animal Species**	**Total number of samples dispatched**
Poultry – Chicken		Wild game – Blesbuck	
Farmed game - Ostrich		Wild game – Other	
Farmed game - other		Cattle	
Wild game – Zebra		Cattle – Dairy	
Wild game – Springbuck			

2. Details of sender – authorized person

Name: _____

Designation: _____

Address: _____

_____

Telephone number: _____

email address: _____

OFFICIAL STAMP

Comments: _____

_____

Signature: _____ Dispatch date: _____

FOR USE BY THE NML OR APPROVED LABORATORY ONLY

Date received:_____ Condition of sample(s) and comments: _____

_____

Signature: _____ Date: _____

ANNEX D: STANDARD OPERATING PROCEDURE TO BE FOLLOWED IN THE CASE OF NON-COMPLIANT SAMPLE RESULTS

1. Introduction

1.1 This standard operating procedure (SOP) lays down rules on specific requirements for official controls and applicable measures for non-compliance or suspected non-compliance with European Union rules applicable to using authorised, unauthorised or prohibited pharmacologically active substances on food-producing animals and to their residues.

2. Definitions

"Unauthorised substances": means pharmacologically active substances, as listed in annex-G and their classification regarding maximum residue limits in food of animal origin or substances that are not authorised to be administered to animals and to be used as feed additives.

"Illegal treatment": means the use in food-producing animals of:

- prohibited or unauthorised substances or products or
- substances or veterinary medicinal products authorised under relevant law for purposes or under conditions other than those laid down in the law.

"Residues of pharmacologically active substances exceeding the maximum residue limit": for the purpose of this SOP, means the presence of residues of authorised pharmacologically active substances in products of animal origin in a concentration exceeding the maximum residue limits set under EU law.

"Residues of pharmacologically active substances exceeding the maximum level": for the purpose of this SOP, means the presence of residues of pharmacologically active substances in products of animal origin, resulting from the unavoidable carry-over of these substances in non-target feed, in a concentration exceeding the maximum levels established by EU legislation and other applicable markets.

3. Responsibility of residue laboratory

3.1. The analyst must immediately inform the laboratory manager of the non-compliant finding. All relevant details pertaining to the sample(s) must be provided.

3.2. The laboratory manager must immediately inform the national executive officer of the non-compliant result(s) by providing an official sample report. All relevant details pertaining to the sample(s) must be provided. A recommendation must be included as to whether a retest or other confirmatory steps are advisable before the matter is accepted as a fact and the official follow-up procedure is activated.

4. Responsibility of National Executive Officer

4.1. The national executive officer (NEO) must inform the provincial executive officer of the non-compliant result(s) by means of a copy of the official laboratory report. All relevant details pertaining to the sample(s) will be provided. Where applicable, copies must also be forwarded to the official veterinarian responsible for the establishment/farm where the sample was collected since non-compliant results may influence his/her decision regarding export certification.

4.2. The NEO must consider the final report made by the provincial executive officer pertaining to the investigation and take any remedial actions necessary to prevent non-conformances of this nature. The National Residue Control Plan may be amended if required.

5. Responsibility of the Provincial Executive Officer

5.1. The provincial executive officer (PEO) in the province must lodge an investigation into the reasons for the non-compliant result(s).

5.2. The following must be included as part of the investigation:

a. Identify the farm of origin.
b. Identify any cohorts of the animal(s) from which the non-compliant sample(s) was (were) collected.
c. Identify any feed, feed additives, water sources, pastures or medication provided or administered to the animal or group of animals and its cohorts from which the non-compliant sample(s) was(were) collected.
d. Keep concerned animal(s) separated from other cohorts. Collect samples from cohorts, other animals under the same production or related circumstances, feed, feed additives, water sources, pasture and any other samples indicated by the circumstances or findings.

5.3. Particular consideration and further (follow-up) investigation must be given to the possible source of the residue or contaminant. Official controls may also include controls on distributors, transporters, veterinary medicines manufacturing premises, feed suppliers and any other site concerned by the investigation.

5.4. At the conclusion of the investigation, consideration must be given to all findings, laboratory results and facts of the case.

5.5. A conclusion, where possible, must be reached, which will fall into one of the following categories:

a. Application of illegal veterinary treatment to animals - use of prohibited or unauthorised pharmacologically active substances or feed additives.
b. Failure to comply with instructions regarding administering authorised veterinary medicines to animals (e.g. the withdrawal period).
c. Indication of environmental contamination and possible source.
d. Inconclusive findings.

5.6. After the investigation, the PEO must complete the following actions:

a. Institute penal action against offenders: Depending on the reason(s) for the non-compliance, penal action can range from a warning, suspension or delisting as a registered export establishment/farm.
b. Take steps to prevent any non-compliant animals from being slaughtered for export.
c. Take steps to prevent the distribution of or recall any non-compliant meat/products where applicable. In the interest of food safety, a recall may be implemented at the discretion of the NEO/PEO before an investigation is complete to prevent further distribution of the product.
d. Compile a detailed report to the NEO explaining the findings of the investigation, the conclusions of the investigation, any remedial actions taken, and any penal actions taken, including any recommendations about the food safety risks and the mitigation thereof gleaned from the investigation and measures taken to prevent recurrence.

ANNEX E: STANDARD OPERATING PROCEDURE TO BE FOLLOWED IN THE CASE OF SUSPECT CARCASSES AT THE ESTABLISHMENT

1. Introduction

1.1. The rules laid down in this standard operating procedure (SOP) must ensure a continuation of the follow-up of suspected or established non-compliance with the rules applicable to the residues or use of pharmacologically active substances authorised in veterinary medicinal products or as feed additives or the use of prohibited or unauthorised pharmacologically active substances.

1.2. Where prohibited or unauthorised substances are discovered in the possession of non-authorised persons, thereby creating a suspicion of illegal treatment and a possible impact on food safety, the measures for official detention (unauthorised medicine/substance, animal, carcase, etc.) and investigations must apply.

2. General principles and actions to be followed at the abattoir

2.1 If the official veterinarian (or officially authorised auxiliary) performing official controls in an abattoir suspect or has evidence that animals have been subject to illegal treatment, he/she must ensure that the following actions are taken:

a. Order that the operator keeps the concerned animals separated from other batches of animals present or arriving at the abattoir.
b. Arrange that the animals be slaughtered separately from other batches of animals
c. Order that the operator separates the carcasses, meat, offal, and by-products from the concerned animals to be immediately identified and kept separated and order such products not to be moved, processed, or disposed of without prior veterinary approval.
d. Order and organize samples necessary to detect the presence of prohibited or unauthorised substances or authorised substances.

2.2 If illegal treatment is established, the provincial executive officer must order the operator to dispose of the carcasses, meat, offal, and by-products.

2.3 If the official veterinarian performing official controls in an abattoir, suspects that the animals present in the abattoir have been treated with an authorised veterinary medicinal product but that the withdrawal period has not been observed, he/she must order that the concerned animals be separated from other batches of animals at the abattoir and must also:

a. postpone the slaughter until the withdrawal period has been observed or
b. issue an order to slaughter the animals separately and, pending the outcome of an investigation, order for the carcasses, meat, offal, and by-products from the concerned animals to be immediately identified and kept separated from other products of animal origin.

2.4 The slaughter may only be postponed temporarily, provided the animal welfare legislation is complied with.

2.5 When the slaughter is postponed in accordance with point 2.3(a) above, the withdrawal period must in no circumstances be shorter than as recommended for the veterinary medicinal product.

2.6 Where necessary, based on evidence of regular use in the particular animal production system, the official veterinarian must institute a programme of regular checking for implantations of growth-promoting substances before or after slaughter or both, as the requirement may be.

2.7 The official veterinarian must perform the following actions when dealing with suspect carcasses:

a. The carcasses are detained for secondary inspection.
b. The suspect treatment sites are not to be removed by the meat inspector or anyone else before the official veterinarian has had a chance to examine the lesions that gave rise to the concern.
c. Samples and any other evidence are collected for further investigation that may be required or indicated, e.g., histopathology, residue screening, and photographic evidence.
d. Suspected carcasses are correctly marked and identified.
e. Conduct a traceability exercise to determine the origin of the carcasses.
f. Pay special attention to other carcasses in the same batch to ensure that similar lesions are not present.
g. Obtain relevant information regarding the animals from the owner of the animals or his representative. If necessary, an on-site visit must be conducted at the farm of origin to obtain information on treatment registers, veterinary drug stock registers, etc.

2.8 When examining the possibility of a chemical risk, the official veterinarian must take the following information into account:

a. Identification numbers of the animal/s, carcasses, and farm of origin.
b. Number of carcasses.
c. Nature of lesions noted.
d. Date.

2.9 The official veterinarian may permit the carcasses to be further handled at the establishment, e.g., deboning, packing, chilling, or freezing, provided that this is done separately from any meat approved for export to the European Union or any other applicable market and as long as full marking and identification of the meat as being suspect is maintained. No approval markings may be applied to the meat or its packing until permission is granted by the official veterinarian at the conclusion of his/her investigation.

2.10 Taking into consideration the results of the information obtained from his/her examination of the carcasses, information gained from the owner, information gained from sample results or any other relevant facts of the case, the official veterinarian must render a judgment on the carcasses regarding their safety from a chemical residue point of view. The following principles must apply:

a. If chemical residues are present but within legal limits of South Africa: Approve meat for local consumption.
b. If chemical residues are present but within legal limits of the European Union: Approve meat for export to the European Union.
c. If chemical residues are present but within legal limits of the applicable export market: Approve meat for export to the respective market.
d. If chemical residues are present but not within the legal limits in South Africa, handle the meat as per legislative provisions.
e. If chemical residues are present but exceed the legal limits of the applicable export market, the meat may not be certified for export to the respective market.

3. Training

3.1. The official veterinarian in his routine training activities at the abattoir must include a training module for training of meat inspectors. The module must train the meat inspectors to identify animals and carcasses that have been or could have been treated with or could otherwise have been exposed to chemicals that may cause unacceptable residues to be present in the meat and other products for human consumption (e.g., liver, kidneys).

3.2. The training must include, amongst others, the following aspects:

a. It is important to identify carcasses that may pose a chemical residue risk.
b. Findings that would indicate the treatment of animals before dispatch to the abattoir.
c. Probable/preferred injection sites on carcasses.
d. The pathological appearance of injection or implantation sites on a carcass.
e. Correct primary meat inspection judgment to be taken when suspect carcasses are encountered.
f. The correct procedure to be followed if suspect carcasses are encountered.

3.3. Records of training conducted in previous years should be available, such as an agenda, list of participants, training material, etc., and must be retained for audit purposes.

ANNEX F: STANDARD OPERATING PROCEDURE FOR THE COMPILATION OF THE NATIONAL RESIDUE CONTROL PLAN APPLICABLE TO EUROPEAN UNION-APPROVED/LISTED FARMS/ESTABLISHMENTS AND OTHER APPLICABLE MARKETS

1. The content of the National Residue Control Plan (NRCP) must include the following:

1.1. List of categories of food-producing animals and products of animal origin covered by the control plan, including details on the species and sub-species of animals.

1.2. Information on the origin of the food-producing animals and products of animal origin.

1.3. Pharmacologically active substances Group A and Group B (Annex G):

a. List of groups of substances covered by the control plan.
b. List of substances and their marker residues to be analysed for the specific animal species and products in the specific matrices, including a justification for their selection based on the risk criteria.
c. The number of samples per animal species and product for each group of substances covered by the control plan.
d. Description of the sampling strategy.

1.4. Pesticides

a. List of substances tested for in the residue control plan.
b. The number of samples per animal species and product for each group of substances covered by the control plan.
c. Justification for selecting pesticides covered by the plan, considering the risks from animal feed, the environment, and the pesticides for which MRLs are established.
d. Justification for the number of samples planned, based on the confidence level achieved in identifying a certain percentage of exceeding the MRLs.

1.5. Contaminants

a. List of substances tested for in the control plan.
b. Justification for the selection of substances.
c. Justification for the selection of contaminants covered by the control plan for pharmacologically active substances, pesticides, and contaminants, taking into account the risks from animal feed and the environment and the contaminants for which MLs have been set.

1.6. National production data from the previous year for the animal species and products of animal origin covered by the control plan must be obtained. The data must include:

a. Number of animal carcasses processed per export approved establishment for the preceding calendar year (by species of interest).
b. Number of animal farms in each province (by species of interest).
c. Number of milk processing establishments in each province.
d. Number of beekeepers (apiary) in each province.
e. Number of tonnes of food-producing animal meat exported during the previous year (by species).
f. Carcass throughput numbers for all abattoirs approved for the EU and other applicable markets.
g. Number of litres of milk processed per export approved establishment for the preceding year (cow milk and other).
h. Number of tonnes/kilograms of honey harvested and processed per export approved establishment for the preceding year.
i. Number of eggs collected/processed per export approved establishment for the preceding year (by species of interest).

1.7. An explanation of whether, for the animals and products of animal origin concerned, the control plan covers the total national production or a proportion of the national production (for example, the production of certain farms/establishments intended for entry into the EU- segregated or split system has been in place).

1.8. If only part of the national production is covered, a description of the system in place to ensure that only those animals and products of animal origin from that segregated population covered by the control plan are eligible for entry into the EU and, where applicable, other respective markets

2. The existing/current NRCP must be saved as a new copy to serve as the basis for compiling a revised plan.

3. The data obtained in paragraph 1 above must be compared with the data from the previous year that was used to compile the current plan. If the data remained relatively similar or decreased in quantity, the number of samples collected must stay the same for the new plan. If the data indicates a general increase, sampling must be increased pro-rata to reflect the increased activity.

4. The previous year's NRCP results must be evaluated and considered valuable risk criteria for the subsequent NRCP drafting, especially non-compliant and suspect samples/results recorded.

5. Results of the current and previous year's National Residue Monitoring Plan (NRMP) must be evaluated and considered as valuable risk criteria for the subsequent NRCP drafting, especially non-compliant and suspect samples/results recorded, considering follow-up action information.

6. The sample groups, substances, and their active metabolites (matrix), type of samples, analysis methods, MRLs and MLs must remain unchanged unless these have changed in the EU, international standards or local legislation since the current plan was implemented. The outcome of the annual risk assessment envisaged in Part III of VPN 19 must also be considered.

7. As soon as the new plan is completed, it must be translated into Sample Collection Grids to be issued per farm/establishment in each province. In compiling the Sample Collection Grids, the following principles must be adhered to:

7.1 The number of samples collected from each farm/establishment must be according to the pro-rata contribution to the total export data during the previous year according to the data obtained under point 1 above.

7.2 Samples must be collected equally throughout the year with the proviso that natural production/processing cycles may be considered.

7.3 Sample collection must be discontinued to provide enough time for the laboratories to complete the analysis and submit the last results to the national executive officer by 28 February each year.

7.4 The number of samples reflected in the Sample Collection Grids must not exceed the capacity of the laboratories to perform the analysis in an appropriate and reasonable time.

8. The NRCP, the revised VPN 19, and the Sample Collection Grids must be subjected to a review to ensure that it reflects all of the required information set out in Annex II to Regulation (EU) 2022/2292 (third country control plan for pharmacologically active substances, pesticides, and contaminants).

9. As soon as the NRCP is approved, the plan must be circulated to the National Mandated Laboratory (NML) and/or the EU-accredited Laboratory for comment. The Sample Collection Grids for the new sampling year must be completed by 30 November each year.

10. Before the circulation of the documentation mentioned in point 9 above, the national executive officer must arrange a meeting with the NML to discuss, amongst others:

    10.1 The practicality of the proposed plan.

    10.2 Any constraints that may be envisaged for the coming year.

    10.3 Any work contracted out to other laboratories and the implications and time frames thereof.

    10.4 The budget requirements that the national executive officer must provide to the NML.

    10.5 Whether it will be possible to maintain reasonable turnover times from sample arrival to sample reports.

    10.6 What planning/procedures are in place to ensure that the role of the NML as stipulated in Regulation (EU) 2017/625 must be complied with during the sampling year (Organise inter-laboratory comparative testing or proficiency tests between official laboratories and ensuring follow-up of such comparative testing or proficiency. Ensure the dissemination to the competent authorities and official laboratories of information that the NML supplies).

11. Considering the comments received from NML, the NRCP and the Sample Collection Grids must be completed.

12. The completed NRCP must be sent to the European Commission and other relevant trading partners upon request on or before 31 March each year (for the EU). It must be accompanied by the completed report for the NRCP of the previous year, completed on the EU-prescribed template.

13. A cover letter must be compiled to accompany the NRCP to the Commission (refer to point 12 above). The letter must state clearly the most significant changes that have been made to the NRCP since the previous one that was submitted. Where required, Table 1, required by the EU, must be completed and attached.

14. Cognition must be given, and amendments must be made, if necessary, to any comments received from the EU and other relevant trading partners after receipt of the NRCP.

15. The NRCP must be implemented from 1 January to 31 December each year.

16. The collection of samples in the provinces must be regularly monitored by the national executive officer, at least quarterly.

17. The NML must be audited by the national executive officer regularly, but at least annually. The purpose of this audit is to verify that all the samples are received, registered, and analysed in a timely manner according to the proposed plan.

18. Review of the NRCP and the Sample Collection Grids.

    The NRCP VPN 19 and the Sample Collection Grids must respond to the following:

    18.1. Are the implemented requirements applicable to South Africa controls on residues of pharmacologically active substances (in veterinary medicinal products), pesticides and contaminants in food-producing animals and animal products for human consumption and intended for the EU market (previously set out in Council Directive 96/23/EC) and repealed by Regulation (EU) 2022/2292 with regard to requirements for the entry into the EU of consignments of food-producing animals and certain goods intended for human consumption?

    18.2. Are all substance groups and substances required by the EU included in the plan? (Requirements for residues of pharmacologically active substances in animals and

food derived therefrom, official controls carried out by competent authorities and annual control plans required by Regulation (EU) 2022/1644 and the arrangements for performing these laid down in Regulation (EU) 2022/1646)?

18.3. Are the required controls on residues of pesticides and contaminants in listed commodities implemented? (In accordance with Regulation (EU) 2021/1355 on multi-annual national control programmes for pesticide residues and Regulation (EU) 2022/932 on uniform practical arrangements for the performance of official controls as regards contaminants in food, on specific additional content of multi-annual national control plans and specific additional arrangements for their preparation)?

18.4. Are the MRLs in the plan in line with the EU requirements? (Regulation (EC) No 470/2009 laying down European Community procedures for establishing residue limits of pharmacologically active substances in foodstuffs of animal origin)?

18.5. Does the number of samples in the plan comply with the EU requirements? (Part II of Annex I to Regulation (EU) 2022/2292)?

18.6. Are the sample methods and sample identification clearly detailed?

18.7. Is there a list of approved Laboratories available? Have the laboratories been approved in writing, and does a valid service delivery agreement exist for each laboratory?

18.8. Has the NML scheduled audit dates of the contract laboratories? Is an audit checklist available? Are any reports available?

18.9. Is the sampling method of live animals adequately described?

18.10. Is the sampling method of feed samples adequately described?

18.11. Do the prescriptions for sample collection include clear instructions for sampling as many different farms as possible?

18.12. Will turnaround times at the laboratories be acceptable?

18.13. Has the principle of sampling without advanced notification been incorporated and clearly communicated?

18.14. Has a clear procedure been incorporated in VPN 19 to guide the investigation of non-compliant results that are obtained at the laboratory?

18.15. Has a clear procedure been incorporated in VPN 19 to guide the punitive measures that must be taken in cases where non-compliant results are obtained at the laboratory?

18.16. Are there clear procedures compiled for the official veterinarian/Authorised Person at an establishment to follow in cases where he/she suspects that animals/carcasses presented for processing may be contaminated with residues?

19. An appropriate, systematized risk analysis must be followed.

ANNEX G: LIST OF PHARMACOLOGICALLY ACTIVE SUBSTANCES

Group A – Prohibited or unauthorised pharmacologically active substances in food-producing animals

1. Substances with hormonal and thyrostatic action and beta-agonists, the use of which is prohibited under Council Directive 96/22/EC [(1)] as amended:
    (a) Stilbenes
    (b) Antithyroid agents
    (c) Steroids
    (d) Resorcylic acid lactones, including zeranol
    (e) Beta-agonists.

2. Prohibited substances listed in Table 2 of the Annex to Regulation (EU) No. 37/2010:
    (a) Chloramphenicol
    (b) Nitrofurans
    (c) Dimetridazole, metronidazole, ronidazole and other nitro-imidazoles
    (d) Other substances

3. Pharmacologically active substances not listed in Table 1 of the Annex to Regulation (EU) No 37/2010[(2)] or substances not authorised for use in feed for food-producing animals in the European Union (EU) according to Regulation (EU) No 1831/2003 of the European Parliament and of the Council [(3)]:
    (a) Dyes
    (b) Plant protection products as defined in Regulation (EU) No 1107/2009 of the European Parliament and of the Council [(4)] and biocides as defined in Regulation (EU) No 528/2012 of the European Parliament and of the Council [(5)] which may be used in animal husbandry of food-producing animals
    (c) Antimicrobial substances.
    (d) Coccidiostats, histomonostats and other antiparasitic agents.
    (e) Protein and peptide hormones
    (f) Anti-inflammatory substances, sedatives, and any other pharmacologically active substances
    (g) Antiviral substances

Group B – Pharmacologically active substances authorised for use in food-producing animals

1. Pharmacologically active substances listed in Table 1 of the Annex to Regulation (EU) No 37/2010:
    (a) Antimicrobial substances
    (b) Insecticides, fungicides, anthelmintics and other antiparasitic agents
    (c) Sedatives
    (d) Non-steroidal anti-inflammatory drugs (NSAIDs), corticosteroids and glucocorticoids
    (e) Other pharmacologically active substances.

2. Coccidiostats and histomonostats authorised according to EU legislation, for which maximum levels and maximum residue limits are set under EU legislation.

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(1) Council Directive 96/22/EC of 29 April 1996 concerning the prohibition on the use in stock farming of certain substances having a hormonal or thyrostatic action and of ß-agonists, and repealing Directives 81/602/EEC, 88/146/EEC, and 88/299/EEC

(2) Commission Regulation (EU) No 37/2010 of 22 December 2009 on pharmacologically active substances and their classification regarding maximum residue limits in foodstuffs of animal origin

(3) Regulation (EC) No 1831/2003 of the European Parliament and of the Council of 22 September 2003 on additives for use in animal nutrition

(4) Regulation (EC) No 1107/2009 of the European Parliament and of the Council of 21 October 2009 concerning the placing of plant protection products on the market and repealing Council Directives 79/117/EEC and 91/414/EEC

(5) Regulation (EU) No 528/2012 of the European Parliament and of the Council of 22 May 2012 concerning the making available on the market and use of biocidal products

ANNEX H: STANDARD OPERATING PROCEDURE FOR OSTRICH AND WILD GAME SAMPLING

1. INTRODUCTION

1.1. South Africa is listed in the Annex to Commission Implementing Regulation (EU) 2021/405 laying down the lists of third countries or regions thereof authorised for the entry into the European Union (EU) of certain animals and goods intended for human consumption in accordance with Regulation (EU) 2017/625 of the European Parliament and the Council as having a residue monitoring plan approved in accordance with Directive 96/23/EC for farmed and wild game (heat-processed ostrich meat, fresh zebra meat and crocodile meat).

1.2. This standard operating procedure has been developed to ensure equivalence with ostrich and wild game sampling within the National Residue Control Plan and relevant EU legislative procedures.

1.3. Blood/serum and feed samples for wild game must not be collected at the farm level due to the nature of wild game not being fed any rations and no treatment administered. Samples of wild game are focused primarily on the abattoir level.

2. COLLECTION OF OSTRICH SERUM SAMPLES

2.1. To ensure compliance with the veterinary residue and food safety legislation of the EU and other relevant markets, it is required to collect serum samples of ostriches at primary production level to test for the presence of prohibited substances and to ensure that other substances are within the prescribed maximum residue levels.

2.2. To comply with the procedures prescribed in this VPN, 2 x 7 ml serum must be collected in gel tubes for each ostrich. Care must be taken that there is at least 5 ml serum per tube (measure this with a ruler - 5 cm will represent about 5 ml). Alternatively, 1 x 10ml serum can be collected in a gel tube. Care must be taken that there is at least 8 ml of serum per tube (measure with a ruler — 8 cm will represent about 8 ml). Samples must be collected in gel tubes from two different randomly selected ostriches.

2.3. These samples may be collected during the annual Newcastle disease or Avian Influenza surveillance programmes. Only Authorised Persons can collect the annual on-farm residue control samples. As practical as possible, farmers must never be informed in advance of the planned collection of serum for residue samples.

3. PROCEDURE FOR SERUM SAMPLE COLLECTION

3.1. Ostriches in the 5 – 14-month groups must be sampled.

3.2. Two tubes of serum (7ml gel tubes) must be collected from each ostrich. Alternatively, one tube (10ml gel tube) may be collected from each ostrich.

3.3. Every serum sample collected must be placed upright in a polystyrene test tube tray.

3.4. The sample submission form (Annex B (1)) must be completed in its entirety and put in an envelope provided by the provincial office. The sample submission form in the envelope must be placed in a plastic bag and transported with the sample. The provincial executive officer must provide all the packaging materials. Where submission forms are incorrectly completed and/or only partially completed, the sample must be rejected for analysis.

3.5. The export registration number of the farm and the sample number of the bird must be written on the tube(s.)

3.6. The bird's identification number (tag number) must be written in the column on the sample submission form (Annex B (1)) to correspond with the sample number written on the tubes and in the column on the sample submission form itself.

3.7. The sex of each bird, where possible to determine, must also be indicated on the tubes.

3.8. The sample submission form (Annex B (1)) provided must be completed for each individual farm (therefore, the sample information of both birds must be on the same form). Where submission forms are incorrectly completed and/or only partially completed, the sample must be rejected for analysis.

3.9. This form must be completed in full.

3.10. The authorised person must be an official veterinarian or an animal health technician.

3.11. The owner, person in charge, or any designated person on the farm must sign the declaration.

3.12. The “Origin of animal” must be completed in full (name of owner, farm name and registration number) and not the farm registration number only. It should be noted that samples whose farm registration is not recorded will not be suitable for analysis and need not even be sent to the NML or EU-accredited laboratory.

3.13. Blood samples must be centrifuged to obtain serum. The serum can then be frozen or chilled (preferably chilled) and forwarded to the national mandated laboratory (NML) or an EU-accredited laboratory as determined by the National Executive Officer (NEO). This procedure must be done under the close supervision of the authorised person.

4. PROCEDURE FOR SERUM SAMPLE DISPATCH

4.1. All samples collected must be dispatched within one week after the collection date. Any samples received later than this period must be rejected for analysis.

4.2. For each batch of samples dispatched, the sample dispatch form (Annex C (3)) must be completed, put in a zip lock bag to protect it from soiling and accompany the container of samples to the NML or EU-accredited laboratory.

4.3. All samples must be packed in an upright position.

4.4. For the dispatch of frozen or chilled samples, a provision must be made to ensure that the samples remain frozen or chilled up to delivery to the NML or EU-accredited laboratory. Note: All samples must be frozen before they are packed into the cooler boxes. Samples do not freeze when the cooler box is placed in the freezer. Samples that are not frozen before they are packed usually arrive rotten at the laboratory.

4.5. The provincial executive officer must supply all the packaging material.

4.6. All residue samples must be couriered directly to the NML or as otherwise directed by the NEO.

4.7. The details of the courier service, where the cost for the service is to be paid for by the competent authority, must be obtained from the NEO.

4.8. Arrangements must be made with the allocated courier service to collect the package for overnight transport to the NML or an otherwise directed location where samples are sent to an EU-accredited laboratory.

4.9. Tamper-proof packaging material must be used for transporting samples, and in case of airfreight, the packaging must comply with the International Air Transport Association (IATA) standards.

4.10. Before dispatch the sample submission form (Annex B (1)) as well as the sample dispatch form (Annex C (3)) must be scanned into electronic copies. The electronic copies must be e-mailed to the provincial executive officer and the national executive officer on the same day the samples are dispatched.

4.11. It is important that a list of farms sampled during the collection year be maintained by the provincial authorities to ensure that all registered farms, as indicated on the sample grids for any particular year, must be sampled and that no repeat sampling must occur.

4.12. A feedback report form must be forwarded from the NML/EU-accredited laboratory to the provincial executive officer, who must forward it to the authorised person who dispatched the samples. This form must be forwarded to confirm receipt of samples and must indicate any non-compliance pertaining to the sampling instructions in this VPN. Prompt corrective actions must be implemented to prevent the recurrence of non-compliance during subsequent sampling.

5. COLLECTION AND DISPATCH OF SAMPLES AT THE ABATTOIR OR ESTABLISHMENT

5.1. Every tissue sample collected must be packed individually, and a sample submission form (Annex A) must be attached to each sample. Where submission forms are incorrectly completed and/or only partially completed, the sample must be rejected for analysis.

5.2. The provincial executive officer must provide sample material.

5.3. Samples must be collected in suitable tamper-proof containers to maintain sample integrity and traceability. In particular, containers must prevent substitution, cross-contamination, and degradation. The containers must be officially sealed.

5.4. Samples at abattoirs must be taken from different farms, and even though different substances are tested, they should not all be taken from one farm. However, in cases where a very low slaughter throughput is experienced at an establishment, it must be permissible to collect more than one sample from a single farm, with the provision that these samples be tested for different substance groups. These cases must be reported in writing to ensure that samples are analysed and not discarded because the samples are collected from the same farm.

5.5. The sample submission form (Annex A) must be completed in its entirety and put in an envelope provided by the provincial executive officer. Where submission forms are incorrectly completed and/or only partially completed, the sample must be rejected for analysis.

5.6. A sample grid with analysis reference numbers must be sent to the provincial executive officers and NML or the EU-accredited laboratories. This analysis reference number must be clearly indicated on the submission form and the envelope of each sample collected and dispatched. The sample submission form in the envelope must be placed in a plastic bag and attached to the sample. The sample with the form attached to it must then be packed in a second zip-lock bag.

5.7. Samples where the envelope is frozen to the sample must be discarded without any action taken by the laboratory to try and identify the sample or its sender. If envelopes are frozen with samples, the submission forms become saturated with water and blood while the sample is thawed, and the submission forms are illegible.

5.8. Note: The following information must be written with a permanent marker on the outside of the envelope, and the envelope must be placed in the plastic bag in such a way that the information is visible from the outside without opening the bag:

a. ZA code of the Abattoir e.g., ZA5
b. Matrix (organ sample) included in the package, e.g., Fat
c. Species, e.g., Ostrich
d. Specific, pre-allocated analysis reference number as indicated in the sample grid for the year, e.g., 92-01/17

5.9. Liver, fat, kidney, and muscle samples must be packed in sample bags provided by the provincial executive officer, securely sealed (tamper proof) and frozen to prevent leakage. The sample with the sample submission form must be packed in a second zip-lock bag and forwarded to the NML or the EU-accredited laboratory.

5.10. Muscle samples denote any muscular tissue of the animal. Muscle samples obtained from the diaphragm must be free from peritoneal or pleural membranes and of the required weight (refer to in the table in section 4.5 of the VPN for ostrich and wild game).

5.11. The whole kidney of the animal must be collected. In the case of animals where the weight of both kidneys is less than 150 g, kidneys from more than one animal from the same farm can be pooled until the weight of the sample weight refer to in the table in section 4.5 of the VPN in the case of Ostrich and wild game. Pooled samples must be indicated as such on the submission form, as this impacts how the samples are prepared for analysis.

5.12. Fat must be sampled from the kidney area in the case of animals where fat is not available from the abdominal cavity. It must be free from blood and other tissues. In the case of animals were not enough fat can be collected from one animal to ensure that the minimum weight of the sample refer to in the table in section 4.5 of the VPN, fat from more than one animal from the same farm can be pooled until the weight of the sample is at least 150 g. Pooled samples must be indicated as such on the submission form, as this impacts how the samples are prepared for analysis.

5.13. In the case of animals where the weight of the liver is less than 150 g, livers from more than one animal from the same farm can be pooled until the weight of the sample is at least 150 g. Pooled samples must be indicated as such on the submission form, as this impacts how the samples are prepared for analysis. For other species pooled minimum weight for liver samples refer to in the table in section 4.5 of the VPN.

5.14. Samples collected must be free from faecal contamination or any other foreign material.

5.15. For each batch of samples dispatched, the sample dispatch form (Annex C (1)) must be completed, put in a zip lock bag to protect it from soiling and accompany the samples to the NML or EU-accredited laboratory.

5.16. A feedback report form must be forwarded from the NML or EU-accredited laboratory to the provincial executive officer, who must forward it to the Authorised person who dispatched the samples. This form must be forwarded to confirm receipt of samples and must indicate any non-conformances pertaining to the sampling instructions in this VPN.

5.17. Prompt corrective actions must be implemented to prevent the recurrence of non-compliance during subsequent sampling.

5.18. Collected samples must be dispatched to the laboratory as soon as possible, but no longer than 3 days after collection.

5.19. Before dispatch the sample submission form (Annex A) as well as the sample dispatch form (Annex C (1)) must be scanned into electronic copies. The electronic copies must be e-mailed to the provincial executive officer and the national executive officer on the same day the samples are dispatched.

6. ON-FARM TARGETED SAMPLING

6.1. Criteria for the selection of targeted samples on farms

(1) All registered ostrich export farms indicated on the particular sample grid for the year must be subjected to on-farm sampling., unless determined otherwise by the NRCP. Birds for sampling can be chosen using local knowledge or any other relevant information, such as the type of fattening system, breed, and animal sex. The authorised person then assesses all the stock on the farm to select the animals to be sampled. In making this assessment, the following criteria should, among other things, be applied:

a. indications of use of pharmacologically active substances,
b. secondary sexual characteristics,
c. behavioural changes,

d. the same level of development in a group of animals of different breeds/categories,
e. animals with good conformation and little fat.

6.2. Type of targeted sample to be collected

(1) The corresponding suitable samples are taken according to the provisions in the residue control plan for export to detect pharmacologically active substances.

7. TARGETED SAMPLING AT PRIMARY PROCESSING ESTABLISHMENTS

7.1. In making their assessment of the animal/bird carcasses and/or the animal products to be sampled, the authorised person must apply the following criteria:

a. species and farming system (feedlot or free-range).
b. information about the producer.
c. indication of the use of pharmacologically active substances.
d. Common practice with regard to the administration of particular pharmacologically active substances in the respective farm production system.

7.2. When taking the samples, efforts should be made to avoid multiple sampling from one producer.

7.3. To detect pharmacologically active substances, the corresponding suitable samples are taken according to the provisions in the annually updated residue control plan.

8. COLLECTION OF FEED SAMPLES

8.1. Definitions:

a. Sampled portion — This refers to the total amount of feed present (in a feed trough(s), camp(s), in a collection of feed bags or a bulk feed bin(s)) that must be sampled and that is homogenous.

b. Aggregate sample — This refers to one representative sample comprising a number of smaller samples [called incremental samples] obtained from the sampled portion by drawing various samples.

c. Incremental samples - This is the number of samples that make up the one aggregate sample and must equal the number indicated on the sample form (Annex B), must be collected at random from different representative places in the sampled portion and must all be more or less equal in size.

d. Reduced sample — means the aggregate sample after it has been thoroughly mixed into one homogenous sample.

e. Final sample — a final sample of at least 500 g. is collected from the reduced sample.

8.2. The purpose of sampling feed is to rule out the addition of growth-promoting substances to manufactured feed, whether done purposefully or accidentally. Only feed to which a premix of medication (concentrated powder form) could reasonably have been added must be sampled.

8.3. The presence of growth-promoting substances is not only a risk with commercially manufactured feeds, where it is indeed possible to add these substances to the feed (in a premix format) during manufacturing or where unintentional cross-contamination from previously manufactured batches may occur but may also be added purposefully or accidentally in feeds mixed by the farmer on the farm for own use.

8.4. Feed sampling must not be limited to commercially manufactured feeds but include commercial and farm-mixed feeds. No roughage samples (e.g., only maize, cut

lucerne, etc.) must be collected (to which it is impossible to add any premixes or medication).

8.5. Further, the risk of slaughter birds is of special significance, and manufactured feed for this group must be sampled in particular rather than fed to ostrich chicks or breeder birds. Feed for slaughter birds includes either grower rations or finishing rations. When samples are collected, it is important to include the feed fed to ostriches from which blood is collected in the feed sample if blood is collected during the same visit.

8.6. It is possible that no samples will be collected on some farms because no commercially manufactured or mixed feed is ever fed on the farm. Provinces must create a means of recording these no-sample reports in the notification process stipulated in section 4.10 of this annexure.

8.7. It is further possible that authorised persons will have to make repeated visits to farms to obtain feed samples, especially in cases where commercially manufactured feed or feed mixed for own purposes are only fed at certain times during the year, at certain times during the production cycle or under certain climatological circumstances.

8.8. The provincial executive officer must provide containers to collect the samples.

8.9. Feed samples must be collected in the following way:

a. Apparatus used for sampling feed, e.g., spade, shovel, spear, mixing (reduction) vessel, sample container, etc., must be constructed and cleaned to such an extent that no sample contamination is possible.

b. The method of feed sample collection must preclude any contamination or change of the sample content.

c. Containers must be labelled and sealed so that the label is destroyed if the container is opened.

d. The sample submission form (Annex B (2)) must be completed in its entirety and put in an envelope attached to the feed sample, including it in a bag with the sample.

e. Before dispatch the sample submission form (Annex B (2)) as well as the sample dispatch form (Annex C (2)) must be scanned into electronic copies. The electronic copies must be e-mailed to the provincial executive officer and the national executive officer on the same day the samples are dispatched.

f. Feed samples must be dispatched to the NML or an EU-accredited laboratory as soon as possible after collection, but not later than 3 days after collection.

g. Each sample sent to the NML, or the EU-accredited laboratory must be accompanied by the sample submission form (Annex B (2)).

h. Where submission forms are incorrectly completed and/or only partially completed, the sample must be rejected for analysis.

8.10. If any clarification on the above is needed, it must be directed to the national executive officer and not the laboratory.

9. SAMPLE NUMBERS

1. The minimum sample numbers must be defined in the NRCP applicable for that specific year. A sample grid must be sent to each province and/or collection official, specifying the number of samples and collection frequency for that particular year.

10. SAMPLE SUBMISSION

1. Please refer to Annex A, B (1) and B (2) for the sample submission forms. The national or provincial executive officer must provide original copies of sample submission forms. Each sample must be accompanied by an original submission form, duly completed, signed, and officially stamped. Where submission forms are incorrectly completed

and/or only partially completed, the sample must be rejected for analysis. The authorised person or office responsible for collecting the sample must always keep a copy.

2. No samples are allowed to be sent to the NML in a packing container containing any other samples, e.g., for disease control purposes, etc. The original sample submission form report remains at the testing laboratory, which has to guarantee that unauthorised persons cannot access this original submission.

ANNEX I: STANDARD OPERATING PROCEDURE FOR THE POULTRY SAMPLING

1. INTRODUCTION

1.1. This standard operating procedure has been developed to ensure equivalence with chicken sampling within the National Residue Control Plan and relevant European Union (EU) legislative procedures.

2. PROCEDURE FOR SERUM SAMPLE COLLECTION

2.1. Collecting chicken serum samples at a primary production level is required to test for prohibited substances and hormonal/antimicrobial growth promoters.

2.2. Broiler chickens can be humanely bled from 21 days of age.

2.3. Sampling procedure must comply with legal provisions and requirements related to animal welfare.

2.4. For every epidemiological unit, randomly selected birds per house must be bled. A 4ml gel serum tube must be used.

2.5. Blood must be taken with a sterile needle from the wing vein (max. 2 ml from one bird). Collective samples from at least 25 birds to make a total of 30 ml must be collected from the same flock/production batch.

2.6. The blood should be cooled and stored at a temperature of +2 to +7˚C and the samples should be delivered to the laboratory within 48 hours.

2.7. Birds from which blood samples have been taken must be separated to avoid cannibalism.

2.8. All serum samples must be placed upright in a polystyrene test tube tray.

2.9. The sample submission form (Annex B (1)) must be completed in its entirety and put in an envelope provided by the provincial office. The sample submission form in the envelope must be placed in a plastic bag and transported with the sample. The provincial executive officer must provide all the packaging materials. Where submission forms are incorrectly completed and/or only partially completed, the sample must be rejected for analysis.

2.10. The export registration number of the farm, as well as the number of the specific house on the farm where the sample was collected, must be written on the tube(s).

2.11. The sample submission form (Annex B (1)) provided must be completed for each farm. Where submission forms are incorrectly completed and/or only partially completed, the sample must be rejected for analysis.

2.12. This form must be completed in full.

2.13. The authorised person must be an official veterinarian or animal health technician.

2.14. The owner, person in charge, or any designated person on the farm must sign the declaration.

2.15. The details of the flock of origin must be completed in full (name of owner/ company name, farm name and registration number) and not the farm registration number only. It should be noted that samples whose farm registration is not recorded will not be

suitable for analysis and need not even be sent to the NML or EU-accredited laboratory.

2.16. Blood samples must be centrifuged to obtain serum. Each 4 ml tube gives 1 ml serum. The serum samples per house must be pooled to give pooled samples. The serum can then be frozen or chilled (preferably chilled) and forwarded to the NML or EU-accredited laboratory. This procedure must be done under the close supervision of the authorised person.

2.17. The authorised person must ensure that the samples are stored and transported under controlled conditions specified in laboratory instructions to prevent degradation or contamination

3. PROCEDURE FOR SERUM SAMPLE DISPATCH

3.1. The provincial executive officer must supply all the packaging material.

3.2. Blood should be collected during bleeding into clean tube/vacuum containers. From this tube/container, the blood must be placed in a test tube (gel tube or vacutainer with anticoagulant). A pooled sample must be collected from the same production batch. The first blood flow should not be collected.

3.3. For each batch of samples dispatched, the sample dispatch form (Annex C (3)) must be completed, put in a zip lock bag to protect it from soiling and accompany the container of samples to the NML or EU-accredited laboratory.

3.4. All samples must be packed in an upright position.

3.5. For the dispatch of frozen or chilled samples, provision must be made to ensure that the samples remain frozen or chilled until delivery to the respective laboratory. Note: All samples must be frozen before they are packed into the cooler boxes. Samples do not freeze when the cooler box is placed in the freezer. Samples that are not frozen before they are packed usually arrive rotten at the laboratory.

3.6. The details of the courier service, where the cost for the service is to be paid for by the competent authority, must be obtained from the NEO.

3.7. Arrangements must be made with the allocated courier service to collect the package for overnight transport to the NML or an otherwise directed location where samples are sent to an EU-accredited laboratory.

3.8. Tamper-proof packaging material must be used for transporting samples, and in case of airfreight, the packaging must comply with the International Air Transport Association (IATA) standards.

3.9. Before dispatch the sample submission form (Annex B (1)) as well as the sample dispatch form (Annex C (3)) must be scanned into electronic copies. The electronic copies must be e-mailed to the provincial executive officer and the national executive officer on the same day the samples are dispatched

3.10. All residue samples must be couriered directly to the NML or EU-accredited laboratory.

3.11. All samples collected must be dispatched within 48 hours after the collection date. Any samples received later than this period must be rejected for analysis.

3.12. The provincial executive officer must maintain a list of farms sampled during the collection year to ensure that all registered farms are sampled and that no repeat sampling must occur.

3.13. A feedback report form must be forwarded from the NML or EU-accredited laboratory to the provincial executive officer, who must forward it to the establishment via the authorised person/state veterinarian who dispatched the samples. This form must be forwarded to confirm receipt of samples and must indicate any non-compliance pertaining to the sampling instructions in this VPN. Prompt corrective actions must be implemented to prevent the recurrence of non-compliance during subsequent sampling.

4. COLLECTION AND DISPATCH OF SAMPLES AT THE ABATTOIR

4.1. A sample grid with analysis reference numbers must be sent to the provincial executive officers and the NML or EU-accredited laboratory.

4.2. The provincial executive officer must provide sampling material.

4.3. Each tissue sample collected must be packed individually, and a sample submission form (Annex A) must be attached to each sample. Where submission forms are incorrectly completed and/or only partially completed, the sample must be rejected for analysis.

4.4. Samples must be collected in suitable tamper-proof containers to maintain sample integrity and traceability. In particular, containers must prevent substitution, cross-contamination, and degradation. The containers must be officially sealed.

4.5. Samples at abattoirs must be taken from chickens from different farms, and even though different substances are tested, they should not all be taken from one farm. However, in cases where a very low slaughter throughput is experienced at an establishment, it must be permissible to collect more than one sample from a single farm, with the provision that these samples be tested for different substance groups. These cases must be reported in writing to ensure that samples are analysed and not discarded because the samples are collected from the same farm.

4.6. The sample submission form (Annex A) must be completed in its entirety and put in an envelope provided. Where submission forms are incorrectly completed and/or only partially completed, the sample must be rejected for analysis.

4.7. The analysis reference number must be clearly indicated on the submission form and the envelope of each sample collected and dispatched. The sample submission form in the envelope must be placed in a plastic bag and attached to the sample. The sample with the form attached to it must then be packed in a second zip-lock bag.

4.8. Liver and muscle samples must be packed in tamper-proof sampling bags to prevent leakage. The sample with the sample submission form attached to it must be packed in a second zip-lock bag and forwarded to the NML or EU-accredited laboratory.

4.9. A muscle sample of 100 g must be obtained by pooling breast meat samples of at least 6 birds from the same flock.

4.10. Where the weight of the liver is less than 150 g, livers from more than one bird from the same farm must be pooled until the weight of the sample reaches the weight referred to in the table in Part 4.5 of the VPN. Pooled samples must be indicated as such on the submission form, as this impacts how the samples are prepared for analysis.

4.11. Samples collected must be free from faecal contamination or any other foreign material.

4.12. Samples where the envelope is frozen to the sample must be discarded without any action taken by the laboratory to try and identify the sample or its sender. If envelopes are frozen to samples, the submission forms become saturated with water and blood while the sample is thawed, and the submission forms are then illegible.

4.13. Note: The following information must be written with a permanent marker on the outside of the envelope, and the envelope must be placed in the plastic bag in such a way that the information is visible from the outside without opening the bag:

a. ZA code of the Abattoir e.g., ZA 5
b. Matrix (organ sample) included in the Package, e.g., Fat
c. Species, e.g. Chicken
d. Specific, pre-allocated analysis reference number as indicated in the sample grid for the year. e.g. 92-01/17

4.14. For each batch of samples dispatched, the sample dispatch form (Annex C (1)) must be completed, put in a zip lock bag to protect it from soiling and accompany the container of samples to the laboratory.

4.15. A feedback report form must be forwarded from the NML or EU-accredited laboratory to the provincial executive officer, who must forward it to the Authorised person who dispatched the samples. This form must be forwarded to confirm receipt of samples and must indicate any non-conformances pertaining to the sampling instructions in this VPN.

4.16. Prompt corrective actions must be implemented to prevent non-compliance recurrence during subsequent sampling.

4.17. Collected samples must be dispatched to the laboratory as soon as possible.

4.18. Before dispatch the sample submission form (Annex A) as well as the sample dispatch form (Annex C (1)) must be scanned into electronic copies. The electronic copies must be e-mailed to the provincial executive officer and the national executive officer on the same day the samples are dispatched.

5. ON-FARM TARGETED SAMPLING

5.1 Criteria for the selection of targeted samples on farms

(1) All registered poultry export farms must be subjected to on-farm sampling. Animals for sampling can be chosen using local knowledge or any other relevant information, such as type of fattening system, breeds and sex of the birds. The authorised person then assesses all the birds on the farm to select the ones to be sampled. In making this assessment, the following criteria should be applied, among other things:

a. indication of the use of pharmacologically active substances,
b. signs of disease or chronic disorder,
c. behavioural changes,
d. different levels of development within a flock, and
e. birds with well-developed muscles.

5.2 Types of targeted samples to be collected

(1) The corresponding suitable samples are taken according to the provisions in the residue control plan.

6. TARGET SAMPLING AT PRIMARY PROCESSING ESTABLISHMENT – CRITERIA FOR SELECTION

6.1. In making their assessment of the animal/bird carcasses and/or the animal products to be sampled, the inspector should apply the following criteria:

a. species, and farming system (feedlot or free-range),
b. information about the producer,
c. indication of the use of pharmacologically active substances, and
d. common practice with regard to the administration of particular pharmacologically active substances in the respective farm production system.

6.2. When taking the samples, efforts should be made to avoid multiple sampling from one producer.

6.3. To detect pharmacologically active substances, the corresponding suitable samples are taken according to the provisions in the annually updated residue control plan.

7. COLLECTION OF FEED AND WATER SAMPLES

7.1. Definitions:

a. Sampled portion — means the total amount of feed present (in a feed trough(s), camp(s), in a collection of feed bags or a bulk feed bin(s)) that must be sampled and that is homogenous.

b. Aggregate sample — means one representative sample comprising a number of smaller samples (called Incremental samples) obtained from the sampled portion by drawing various samples.

c. Incremental samples – means the number of samples that make up the one aggregate sample and must equal the number indicated on the sample form (Annex B), must be collected at random from different representative places in the sampled portion and must all be more or less equal in size.

d. Reduced sample — means the aggregate sample after it has been thoroughly mixed into one homogenous sample.

e. Final sample — means a final sample of at least 500 g collected from the reduced sample.

7.2. The purpose of sampling feed is to rule out the addition of growth-promoting substances to manufactured feed, whether this was done purposefully or accidentally. Only feed to which a premix of medication (concentrated powder form) could reasonably have been added must be sampled.

7.3. The presence of growth-promoting substances is not only a risk with commercially manufactured feeds, where it is indeed possible to add these substances to the feed (in a premix format) during manufacturing or where unintentional cross-contamination from previously manufactured batches may occur but may also be added purposefully or accidentally in feeds mixed by the farmer on the farm for own use.

7.4. Feed sampling must not be limited to commercially manufactured feeds but must include commercial and farm-mixed feeds. No roughage samples (e.g. only maize, cut lucerne, etc.) must be collected.

7.5. Further, the risk for slaughter birds is of special significance, and manufactured feed for this group must be sampled in particular rather than feed for ostrich chick or breeder birds. Feed for slaughter birds includes either grower rations or finishing rations. When samples are collected, it is important to include the feed fed to birds from which blood is collected in the feed sample if blood is collected during the same visit.

7.6. It is possible that no samples will be collected on some farms because no commercially manufactured or mixed feed is ever fed on the farm. Provinces must create a means of recording these no-sample reports in the notification process.

7.7. It is further possible that authorised persons will have to make repeated visits to farms to obtain feed samples, especially in cases where commercially manufactured feed or feed mixed for own purposes are only fed at certain times during the year, at certain times during the production cycle or under certain climatological circumstances.

7.8. The provincial executive officer must provide all the packaging materials.

7.9. When collecting feed samples from poultry farms, feed for the same poultry category should be collected from feeding equipment (feeders) at multiple locations and put in a clean sampling container.

7.10. Feed samples must be collected in the following way:

a. Apparatus used for sampling feed, e.g. spade, shovel, spear, mixing (reduction) vessel, sample container, etc., must be constructed and clean to such an extent that no sample contamination is possible.

b. The method of feed sample collection must preclude any contamination or change of the sample content.

c. The sampled amount of feed should be mixed well to homogenize, and then 0.5 kg should be collected using its equipment for sampling (e.g. scoop). Note: In order to prevent cross-contamination, farm equipment should not be used to collect or homogenize a sample.

d. Containers must be labelled and sealed so that the label is destroyed if the container is opened.

e. A sample of feed with a high percentage of moisture should be stored in a refrigerator and delivered as soon as possible to not spoil the sample.

f. The Sample Submission Form (Annex B (2)) must be completed in its entirety and put in an envelope attached to the feed sample, including it in a bag with the sample.

g. Before dispatch the sample submission form (Annex B (2)) as well as the sample dispatch form (Annex C (2)) must be scanned into electronic copies. The electronic copies must be e-mailed to the provincial executive officer and the national executive officer on the same day the samples are dispatched.

h. Feed samples must be dispatched to the NML or EU-accredited laboratory as soon as possible after collection, but not later than 3 days after collection.

i. Each sample sent to the NML or EU-accredited laboratory must be accompanied by the sample submission form (Annex B (2)).

j. Where submission forms are incorrectly completed and/or only partially completed, the sample must be rejected for analysis.

7.11. Drinking water samples must be taken from different locations/drinkers/containers from which the birds drink water. 500 ml must be collected, and the sample must be stored at a temperature of +2 to +7˚C before delivery/dispatch to the laboratory.

7.12. If any clarification on the above is needed, it must be directed to the national executive officer and not the laboratory.

8. SAMPLE NUMBERS

8.1. The minimum sample numbers must be defined in the NRCP applicable for that specific year. A sample grid must be sent to each province and/or collection official, specifying the number of samples and collection frequency for that particular year.

9. SAMPLE SUBMISSION

9.1. Please refer to Annexes A, B (1) and B (2) for examples of the sample submission form. The national executive officer or relevant provincial counterpart must provide original copies of sample submission forms. Each sample must be accompanied by an original submission form, duly completed, signed, and officially stamped. Where submission forms are incorrectly completed and/or only partially completed, the sample must be rejected for analysis. The authorised person or office responsible for collecting the sample must always keep a copy.

9.2. No samples are allowed to be sent to the laboratory in a packing container containing any other samples, e.g., for disease control purposes, etc. The original of the sample submission form report remains at the laboratory, which has to guarantee that unauthorised persons cannot access this original Submission form.